



AN EFFICIENT IODOBENZENE DIACETATE (IBD) CATALYZED TETRAZOLO[1,5-*a*] QUINOLINE INCORPORATED 1,3,4-OXADIAZOLE NUCLEUS: SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL EVALUATION

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Abstract

A facile, convenient and high yielding library of 7-substituted tetrazolo [1,5-*a*]quinoline incorporated 1,3,4-oxadiazole nucleus was reported by the cyclization of the corresponding schiff base derivatives. Various substituted schiff base derivatives *i.e.* (**5a-l**) and (**5m-r**) were obtained by the reaction of various tetrazolo[1,5-*a*]quinoline-4-carbaldehyde with substituted benzohydrazide as well as nicotinohydrazide/isonicotinohydrazide respectively. The targeted substituted 1,3,4-oxadiazole derivatives (**6a-r**) were obtained by cyclization with different cyclising reagents among which iodobenzenediacetate was found to be an extremely good catalyst. The structures of the targeted compounds were confirmed by FT-IR, ¹H NMR, ¹³C NMR and mass spectrometry. All the resultant compounds were evaluated for their *in vitro* antibacterial activity against a panel of pathogenic strains of bacteria and fungi, *in vitro* antioxidant activity by the ferric-reducing antioxidant power method, *in vitro* antitubercular activity against *Mycobacterium tuberculosis* H37Rv strain and for their *in vitro* antimalarial activity against *Plasmodium falciparum*. Many candidates of this new class revealed noticeable activities against first line drugs.

Keywords: Tetrazolo[1,5-*a*]quinoline clubbed 1,3,4-oxadiazole nucleus; antimicrobial activity; iodobenzenediacetate (IBD)

1. Introduction

Heterocyclic compounds are the scaffolds on which pharmacophores are arranged to provide potent and selective drugs[I]. This is especially true for five-membered ring heterocyclic compounds. 1,3,4-oxadiazoles and 1,3,4-thiadiazoles have been used as “privileged” scaffolds to produce substances of interest in broaden therapeutic area.[II-VIII] Tuberculosis has become a serious global health problem as it is one of the leading causes of death amongst infectious diseases. About one-third of the world's population is infected by

Mycobacterium tuberculosis every year and more than 2 million deaths are reported[IX]. The World Health Organization has declared TB to be a 'global emergency' and a recent estimation by WHO showed that within the next 20 years around 30 million people will be infected by *M. tuberculosis*. [X, XI] and new forms of tuberculosis like multidrug resistant tuberculosis (MDR TB) and extensively drug resistant tuberculosis (XDR TB) are found to be emerging as new challenge for medicinal chemists[XII].

Malaria remains a significant worldwide health problem, with serious social and economic consequences in affected countries. Malaria is also a critical diseases in public health around the world. It is caused by five different types of protozoan of the genus *Plasmodium*, but *Plasmodium falciparum* is responsible for most of the critical cases[XIII]. Amongst them, to reduce the variety of resistant parasites, the WHO has suggested the combined formulation of artemisinins with traditional antimalarial drugs such as lumefantrine, amodiaquine and mefloquine, and ACT (Artemisinin Combination Therapy) is currently approved in multiple countries [XIV, XV]. Pharmaceutical chemists are in focussing on creation of library of new analogues of existing drugs or new chemical entities with hope to obtain the agents with potential antitubercular activity as well as antimalarial activity with reduced toxicity and treatment duration.

Quinolines represent an essential group of heterocyclic compounds as they are pivotal skeletons in many biologically active natural products as well as various pharmacologically interesting compounds [XVI-XIX]. Azole class of compounds are gaining importance because of their key property lipophilicity that influence the ability of the drug to reach the target by transmembrane diffusion [XX]. Quinoline incorporated 1,3,4-Oxadiazoles have exhibited wide range of biological activities such as antibacterial [XXI, XXII], antitubercular [XXIII, XXIV], antitumor [XXV, XXVI], antifungal [XXVII, XXVIII], anti-inflammatory [XXIX, XXX], antimalarial [XXXI-XXXIV] activities and are key component of antibiotic furamizole [XXXV] as well as antiretroviral raltegravir [XXXVI].

We thought worthwhile to design and synthesize new hybrid substituted tetrazolo[1,5-*a*]quinoline clubbed 1,3,4-oxadiazoles with substituted phenyl and pyridyl ring systems in a single molecular framework with a hope to obtain the new molecules with enhanced antimycobacterial activity.

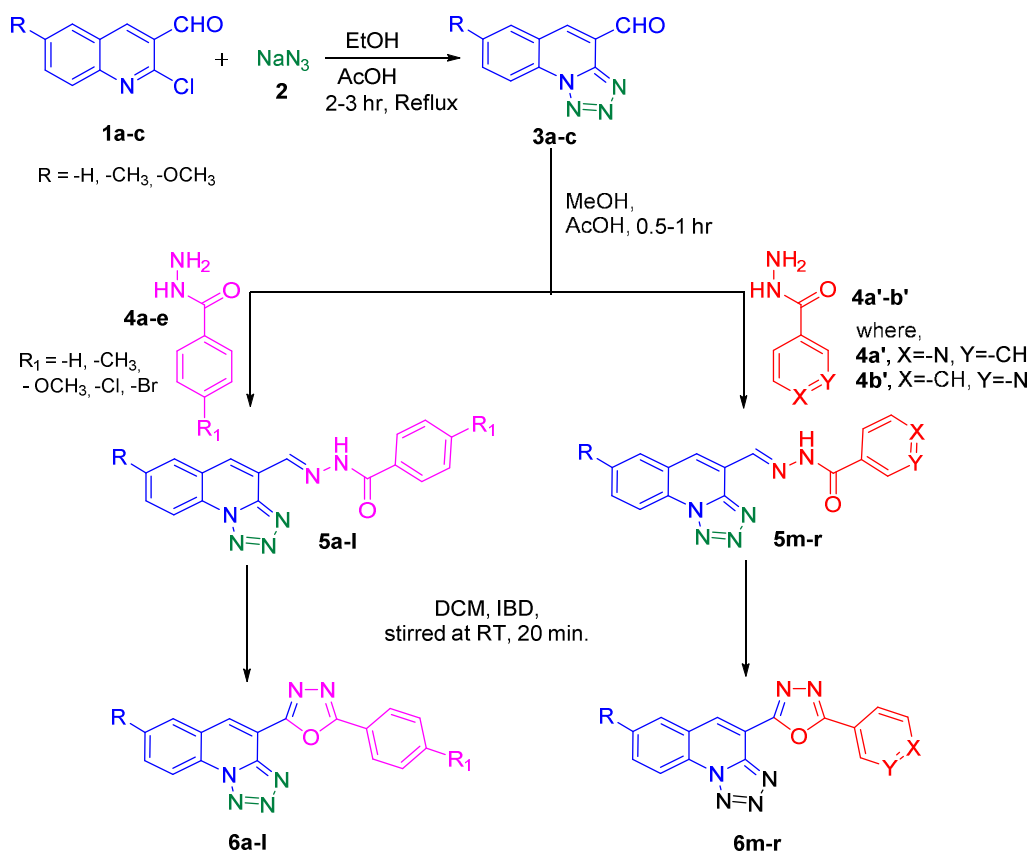
2. Results and discussion

2.1. Chemistry

The synthetic protocol adopted to obtain the targeted quinoline nucleus incorporated 1,3,4-oxadiazole derivatives is depicted in Scheme 1. The starting material 2-chloroquinoline-3-carbaldehydes **1** was prepared according to Vilsmeier-Haack reaction according to a literature procedure [XXXVII]. 7-substituted tetrazolo[1,5-*a*]quinoline-4-carbaldehydes (**3a-c**) were prepared by reacting 7-substituted 2-chloroquinoline-3-carbaldehyde **1** and sodium azide **2** in the presence of catalytic amount of glacial acetic acid and ethanol as a solvent.

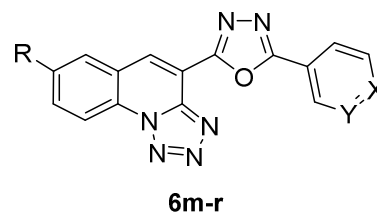
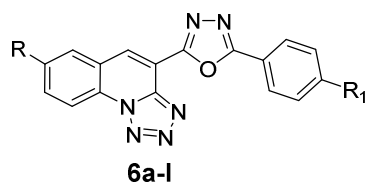
Then after 7-substituted tetrazolo[1,5-*a*]quinoline-4-carbaldehydes (**3a-c**) were refluxed with 4-substitutedbenzohydrazide **4a-e** (10 mmol), nicotinohydrazide (**4a'**) and/or isonicotinohydrazide (**4b'**) in the presence of catalytic amount of glacial acetic acid in methanol (40 mL) was refluxed for 1h to give (E)-4-substituted-N'-((7-substitutedtetrazolo[1,5-*a*]quinolin-4-yl)methylene)benzohydrazide (**5a-l**) and (E)-N'-((7-substitutedtetrazolo[1,5-*a*]quinolin-4-yl)methylene)nicotinohydrazide/isonicotinohydrazide (**5m-r**) respectively. The obtained hydrazones *i.e.* **5a-l** and **5m-r**, were then subjected to an oxidative cyclization using iodobenzenediacetate (IBD) in dichloromethane (DCM) by stirring at room temperature for 20 min to afford corresponding 1,3,4-oxadiazoles *i.e.* **6a-l**

and **6m-r** respectively. The cyclization of the resultant schiff base derivatives *i.e.* **5a-l** and **5m-r** was carried out under different cyclizing reagents such as mercuric acetate, lead dioxide, chloramine-T, iodobenzene diacetate and aqueous sodium hydroxide with iodine in aqueous potassium iodide.



Scheme1. Synthesis of 7-substituted tetrazolo [1,5-a]quinoline incorporating 1,3,4-oxadiazole nucleus (**6a-r**).

The choice of substituents introduced in the synthesized compounds **6a-r** is based on the most influential aspect of lipophilicity as it determines solubility, reactivity and formulation of pharmaceuticals as well as metabolism of drugs.

Table 1. Preliminary characterization of all synthesized compounds **6a-r**.

Comp.	R	R ₁	Yield ^a (%)
6a	H	H	93
6b	H	CH ₃	73
6c	H	OCH ₃	81
6d	H	Cl	80
6e	H	Br	86
6f	CH ₃	H	83
6g	CH ₃	CH ₃	79
6h	CH ₃	OCH ₃	74
6i	CH ₃	Cl	89
6j	OCH ₃	H	76
6k	OCH ₃	CH ₃	84
6l	OCH ₃	OCH ₃	80

Comp.	R	X	Y	Yield ^a (%)
6m	H	N	CH	84
6n	H	CH	N	77
6o	CH ₃	N	CH	86
6p	CH ₃	CH	N	81
6q	OCH ₃	N	CH	85
6r	OCH ₃	CH	N	83

^a Isolated yield

The structures of all the synthesized compounds were confirmed by ¹H NMR, ¹³C NMR, FT-IR, mass spectrometry and elemental analysis. The IR spectrum of all the synthesized compounds showed absorption in the range of 1620–1638 cm⁻¹ due to C=N stretching. The characteristic C–H stretching of aromatic ring was observed at around 3051–3067 cm⁻¹. Strong absorption bands were observed in the range of 1589–1598 cm⁻¹ and 1361–1373 cm⁻¹ due to the presence of the C=C stretching as well as CH₃ group respectively. In the ¹H NMR spectra of the resultant compounds, the aromatic protons of quinoline and benzene resonate as multiplet around δ 7.45–9.31 ppm. The methyl and methoxy protons of the quinoline and benzene ring appeared near to δ 2.33 ppm and δ 3.91 ppm respectively. ¹³C NMR spectra of the 1,3,4-oxadiazole ring, C-2 carbon was displayed as a very downfield signal at δ 164.1–165.9 ppm for the reason that it was shrink between one oxygen atom and one nitrogen atoms. The mass spectra of all the compounds showed molecular ion peaks at M⁺ corresponding to their molecular weights, which confirmed the respective chemical structures.

Scheme 2. Screening of catalysts to optimize reaction condition for the synthesis of tetrazolo [1,5-*a*]quinoline incorporated 1,3,4-oxadiazole **6a**.

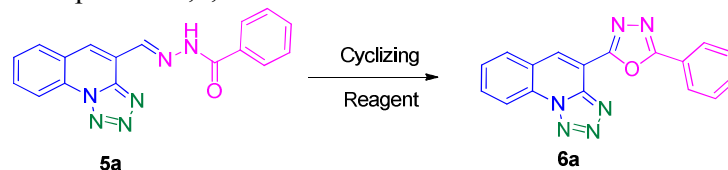


Table 2. The influence of different cyclizing reagents on the model reaction under different conditions

Entry ^a	Catalyst	Solvent	Time ^b (min.)	Yield ^c (%)
1	Conc. H ₂ SO ₄	Acetic acid	50	Trace
2	Chloramine-T	DCM	50	40
3	Hg(OAc) ₂	Methanol	40	45
4	Hg(OAc) ₂	DCM	35	50
5	IBD	DCM	20	93
6	IBD	Methanol	25	70
7	IBD	Ethanol	45	57
8	Aq. NaOH	Methanol	35	47
9	Aq. NaOH	DCM	40	55
10	Aq. NaOH	Ethanol	43	51

^a Reaction of schiff base **5a** (1.0 mmol) in the presence of (1.0 mmol) catalyst.

^b Reaction progress monitored by TLC.

^c Isolated yield.

For the screening of different cyclizing reagents we selected a model reaction between (*E*)-N'-(tetrazolo[1,5-*a*]quinolin-4-ylmethylene)benzohydrazide **5a** and different cyclizing reagents (**Scheme 2**). After cyclization, the corresponding benzohydrazide derivatives get converted into the resultant product *i.e.* 7-substituted tetrazolo [1,5-*a*]quinoline incorporated 1,3,4-oxadiazole **6a-r**. The results are summarized in **Table 2**.

2.2. Optimization of synthetic protocol

2.2.1. Screening of catalysts to optimize the reaction conditions for 7-substituted tetrazolo [1,5-*a*]quinoline incorporated 1,3,4-oxadiazole **6a-r**.

We began our examination with variously substituted schiff base derivatives. Various schiff base derivatives *i.e.* (*E*)-4-substituted-N'-((7-substituted tetrazolo [1,5-*a*]quinolin-4-yl)methylene)benzohydrazide (**5a-l**) and (*E*)-N'-((7-substituted tetrazolo[1,5-*a*]quinolin-4-yl)methylene)nicotinothydrazide/isonicotinothydrazide (**5m-r**) were obtained by the mixing equimolar amounts of tetrazolo[1,5-*a*]quinoline-4-carbaldehyde with the corresponding substituted benzohydrazide / nicotinothydrazide / isonicotinothydrazide in a good to excellent yield in the presence of glacial acetic acid as the catalyst and methanol as a solvent media over a time duration of 0.5-1.0 hr.

To optimize the reaction conditions, a series of experiments were performed with the model reaction of (*E*)-N'-(tetrazolo[1,5-*a*]quinolin-4-ylmethylene)benzohydrazide (1.0 mmol) **5a** and different cyclizing reagents (1.0 mmol) (**Scheme 2**) in the various reaction media.

In order to search for the best solvent, the model reaction was performed in three different solvents such as acetic acid, ethanol, methanol and dichloromethane (DCM) for the

synthesis of **6a**. The results are summarized in **Table 2**. For optimization of the reaction conditions, initially the reaction in acetic acid in the presence of concentrated H₂SO₄ as the catalyst afforded the desired product **6a** in a very trace amount (Table 2, entry 1). After further investigation of various catalysts, IBD was found to be more effective than other catalysts such as chloramine-T, mercuric acetate, Aq. NaOH. IBD showed advantages in the DCM reaction media over the other solvents not only in influencing the reaction but also miscibility of iodobenzene diacetate (IBD) in it and easy separation of the targeted compound to yield **6a** (93%) (Table 2, entry 5).

Due to these results, IBD was selected as the cyclizing reagent for the synthesis of tetrazolo [1,5-*a*]quinoline incorporated 1,3,4-oxadiazole, and thus the reaction was carried out in a single step by stirring different schiff bases **5a-r** in the presence of iodobenzenediacetate (IBD) at room temperature in DCM as a solvent to produce the desired 7-substituted tetrazolo [1,5-*a*]quinoline incorporated 1,3,4-oxadiazole (**6a-r**).

2.3. Pharmacology

2.3.1. *In vitro* antimicrobial activity

Table 3. *In vitro* antimicrobial activity (MIC, µg/mL) of compounds **6a-r**.

Compound	<i>Gram positive bacteria</i>			<i>Gram negative bacteria</i>			<i>Fungi</i>	
	S.P.	B.S.	C.T.	E.C.	S.T.	V.C.	C.A.	A.F.
	MTCC	MTCC	MTCC	MTCC	MTCC	MTCC	MTCC	MTCC
	1936	441	449	443	98	3906	227	3008
6a	500	200	500	100	250	250	>1000	500
6b	100	500	250	200	500	200	1000	>1000
6c	100	200	500	250	200	200	1000	500
6d	62.5	500	1000	250	200	500	250	500
6e	500	250	500	100	250	1000	500	100
6f	500	500	500	250	100	62.5	1000	>1000
6g	100	100	500	500	200	500	1000	1000
6h	250	200	100	100	500	500	500	>1000
6i	200	100	62.5	500	100	100	500	100
6j	500	500	250	100	200	200	1000	1000
6k	100	1000	100	200	200	100	250	1000
6l	500	500	250	500	500	500	1000	100
6m	200	100	100	62.5	100	500	1000	1000
6n	500	62.5	500	100	200	500	250	1000
6o	100	1000	62.5	250	100	100	500	500
6p	500	200	500	500	62.5	100	1000	100
6q	100	200	200	62.5	100	1000	250	500
6r	62.5	1000	100	500	500	100	1000	500
A	100	250	250	100	100	100	n. t. ^a	n. t.
B	10	100	50	10	10	10	n. t.	n. t.
C	50	50	50	50	50	50	n. t.	n. t.
D	25	50	100	25	25	25	n. t.	n. t.
E	n. t.	n. t.	n. t.	n. t.	n. t.	n. t.	100	100
F	n. t.	n. t.	n. t.	n. t.	n. t.	n. t.	500	100

S.P.: *Streptococcus pneumoniae*, B.S.: *Bacillus subtilis*, C.T.: *Clostridium tetani*, E.C.: *Escherichia coli* S.T.: *Salmonella typhi*, V.C.: *Vibrio cholerae*, C.A.: *Candida albicans*, A.F.: *Aspergillus fumigatus*, MTCC: Microbial Type Culture Collection. A: Ampicillin, B:

Norfloxacin, C: Chloramphenicol, D: Ciprofloxacin, E: Nystatin, F: Griseofulvin, ^a n.t.: not tested.

The *in vitro* antimicrobial screening of targeted compounds **6a-r** at minimal inhibitory concentration (MIC) was carried out using the broth microdilution method according to the National Committee for Clinical Laboratory Standards (NCCLS)[XXXVIII, XXXIX]. The antibacterial activity was tested against three Gram positive (*Streptococcus pneumoniae* MTCC 1936, *Bacillus subtilis* MTCC 441 and *Clostridium tetani* MTCC 449) and three Gram negative (*Escherichia coli* MTCC 443, *Salmonella typhi* MTCC 98, and *Vibrio cholera* MTCC 3906) bacteria using ampicillin, ciprofloxacin, norfloxacin and chloramphenicol as the reference antibacterial drugs.

The antifungal activity was screened against two fungal species (*Candida albicans* MTCC 227 and *Aspergillus fumigatus* MTCC 3008) where nystatin and griseofulvin were used as the reference antifungal agents. The strains employed for the activity screening were obtained from the Institute of Microbial Technology, Chandigarh (MTCC – Micro Type Culture Collection). Mueller Hinton broth was used as the nutrient medium to grow and dilute the drug suspension for the test. The results of the antimicrobial screening are shown in **Table 3**.

Upon investigation of Antibacterial activity data (Table 3), revealed that all the compounds **6a-r** showed moderate to very good inhibitory activity. It has been observed that against *S. pneumonia*, compounds **6d** and **6r** were found to be more potent *i.e.* MIC, 62.5 µg/mL as compared to ampicillin *i.e.* 100 µg/mL. Compound **6n** was found to possess excellent potency *i.e.* 62.5 µg/mL as compared to ampicillin *i.e.* 250 µg/mL as well as norfloxacin *i.e.* 100 µg/mL.

Compounds **6g**, **6i** and **6m** were found to be more potent against *B. subtilis* *i.e.* 100 µg/mL as compared to ampicillin. Compounds **6a**, **6c**, **6h**, **6p** and **6q** were found to be more effective (MIC = 200 µg/mL) against *B. subtilis* as compared to ampicillin. Compound **6e** was found equipotent as that of ampicillin against *B. subtilis*. Against *S. pneumonia* compounds **6b**, **6c**, **6g**, **6k** and **6q** showed comparable activity to that of ampicillin. Against *C. tetani*, compounds **6i** and **6o** were found to be more potent *i.e.* MIC, 62.5 µg/mL as compared to ampicillin *i.e.* 100 µg/mL. Against *C. tetani*, compounds **6b**, **6j** and **6l** displayed same influence as that of ampicillin.

In case of gram negative bacteria, it has been observed that against *E. Coli*, compounds **6m** and **6q** were found to possess excellent potency *i.e.* MIC 62.5 µg/mL as compared to ampicillin *i.e.* 100 µg/mL and compound **6p** was found to be more potent *i.e.* MIC 62.5 µg/mL against *S. Typhi* as compared to ampicillin. Against *E. Coli*, compounds **6a**, **6e**, **6h**, **6j** and **6n** were equipotent to that of ampicillin *i.e.* MIC, 100 µg/mL. In case of *S. Typhi*, compounds **6f**, **6i**, **6m**, **6o** and **6q** were found to be equipotent to that of ampicillin *i.e.* 100 µg/mL. Against *V. Cholerae*, compound **6f** was found to have maximum potency *i.e.* 62.5 µg/mL as compared to ampicillin *i.e.* 100 µg/mL. Compounds **6i**, **6k** and **6r** were found equipotent to that of ampicillin *i.e.* 100 µg/mL. Remaining other compounds are moderate or less active against all gram positive and gram negative bacteria.

The result of antifungal study (Table 3) of the synthesized 1,3,4-Oxadiazole derivatives revealed that all the compounds have excellent activity against *A. Fumigates*. The antifungal screening data from Table 2 revealed that against *C. albicans*, compound **6d**, **6k**, **6n** and **6q** *i.e.* 250 µg/mL, were found to be foremost active as compared to griseofulvin. Also against *C. albicans* compounds **6e**, **6h**, **6i** and **6o** showed comparatively similar potency to that of griseofulvin. Compounds **6e**, **6i**, **6l** and **6p** exhibited comparable potency *i.e.* 100 µg/mL against *A. fumigates*. From the above results, we concluded that compound **6q** exhibit very good potency to become new member of antimicrobial agent.

2.3.2. *In vitro* antimalarial activity

All the compounds (**6a-r**) were screened for their *in vitro* antimalarial activity against the *P. falciparum* strain using chloroquine and quinine as the reference compounds. All experiments were performed in duplicate and the results of the antimalarial screening are expressed as the drug concentration resulting in 50% inhibition (IC₅₀) of parasite growth and are listed in **Table 4**.

Table 4. *In vitro* antimalarial activity of compounds **6a-r**.

Compound	IC ₅₀ (µg/mL)	Compound	IC ₅₀ (µg/mL)
6a	0.083	6k	0.82
6b	0.064	6l	1.89
6c	1.55	6m	0.082
6d	0.032	6n	0.85
6e	0.41	6o	1.32
6f	0.78	6p	0.034
6g	1.53	6q	0.073
6h	0.061	6r	0.37
6i	0.58	Chloroquine	0.020
6j	0.51	Quinine	0.268

As shown in **Table 4**, the compounds **6a**, **6b**, **6d**, **6h**, **6m**, **6p** and **6q** were found to have IC₅₀ in the range of 0.032-0.083 against *P. falciparum* strain. These compounds displayed superior activity against *P. falciparum* strain as compared to quinine IC₅₀ 0.268. Moreover compounds **6d** and **6p** were found to possess moderate activity i.e. IC₅₀ 0.032 and 0.034 respectively aligned with chloroquine. Remaining all other compounds were found to be less active against *P. falciparum* strain against chloroquine and quinine as the standard drugs.

2.3.3. *In vitro* antituberculosis activity

The encouraging results from the antibacterial activity screening provoked us to carry out preliminary screening of the title compounds for their *in vitro* antituberculosis activity.

Table 5. *In vitro* antituberculosis activity (% inhibition) of compounds **6a-r** against *M. tuberculosis* H37Rv (at concentration 250 and 100 µg/mL).

Compound	% Inhibition		Compound	% Inhibition	
	250 µg/mL	100 µg/mL		250 µg/mL	100 µg/mL
6a	65	31	6k	78	49
6b	79	54	6l	83	67
6c	35	12	6m	94	69
6d	92	79	6n	84	80
6e	64	43	6o	81	55
6f	48	23	6p	65	45
6g	53	29	6q	98	96
6h	95	94	6r	67	44
6i	87	81	Rifampicin	98	98
6j	71	51	Isoniazid	99	99

Primary screening of targeted compounds **6a-r** were performed at 250 µg/mL against the *Mycobacterium tuberculosis* H37Rv strain using Lowenstein–Jensen medium as described

by Rattan[XL, XLI]. The acquired results are presented in Table 5 in the form of % inhibition. Rifampicin and isoniazid were used as the reference drugs. From these results the five compounds (**6d**, **6h**, **6m**, **6n** and **6q**) that exhibited the highest % inhibition were screened again to get their MIC values (Table 5).

Antituberculosis screening of all the newly synthesized compounds **6a-r** were conducted at two concentrations i.e. 250 µg/mL and 100 µg/mL against tuberculosis H37Rv strain. The bioassay results obtained for the efficacy of all the synthesized analogues against *Mycobacterium tuberculosis* H37Rv is listed in Table 5.

The outcome of the result revealed that, compounds **6d** and **6m** were found to possess excellent activity (i.e. 92% and 94% at 250 mg/mL) against *M. tuberculosis* H37Rv. At the commencement of this study in the preliminary screening, compound **6h** displayed better activity and showed 95% inhibition at 250 µg/mL concentration and 94% inhibition at 100 µg/mL concentration as well as compound **6q** showed better activity and showed 98% inhibition at 250 µg/mL concentration and 96% inhibition at 100 µg/mL concentration. While compounds **6i** and **6n** are moderately active against *M. tuberculosis* H37Rv. All other compounds showed poor inhibition of *M. tuberculosis* growth. From the above results, it can be concluded that, compound **6q** may become new potent member of antitubercular agents in future.

2.3.4. In vitro antioxidant activity

The antioxidant activity of the entire series was investigated by using ascorbic acid as the standard antioxidant compound and results are listed in Table 6. Ferric reducing antioxidant power (FRAP) was measured by a modified method[XLII]. The antioxidant potentials of the compounds were estimated as their power to reduce the TPTZ–Fe(III) complex to the TPTZ–Fe(II) complex. The absorbance of the intense blue coloured [Fe(II)–TPTZ] complex was measured at 593 nm.

Table 6. In vitro antioxidant activity of compounds **6a-r**.

Comp.	OD (593 nm)	FRAP value ^a	Comp.	OD (593 nm)	FRAP value ^a
6a	0.314	203.20	6k	1.163	259.73
6b	1.689	257.18	6l	1.103	220.24
6c	1.993	79.13	6m	2.413	497.43
6d	2.503	493.50	6n	1.701	344.82
6e	1.823	332.13	6o	2.273	458.23
6f	1.783	219.81	6p	2.418	440.02
6g	1.953	387.11	6q	1.288	355.68
6h	2.487	495.12	6r	2.295	481.91
6i	1.804	367.19	A.A.	2.501	
6j	1.273	362.38			

A.A. = ascorbic acid, concentration of compounds used = 200 mg/mL, concentration of standard (A.A.) = 176 mg/mL. ^aA.A. mmol per 100 g of the sample. FRAP = ferric reducing antioxidant power

The standard deviation value is shown in terms of ±SD (≤0.75). All the newly synthesized compounds are screened for their antioxidant activity in three sets (n = 3). The results are expressed as ascorbic equivalents (mmol per 100 g of the compound) in table 6.

Upon the antioxidant assay, synthesized compounds **6d**, **6h**, **6m**, **6o**, **6p** and **6r** gave ferric reducing antioxidant power (FRAP) value ranging from 440.02 to 497.43 mmol per 100 g of compounds. This indicates that these compounds are good antioxidants. Compound **6c** showed poor antioxidant power while the remaining other compounds showed moderate

antioxidant activity (Table 6).

2.4. Structure-activity relationship (SAR)

The structure-activity relationship study (Fig. 1) revealed that various substituents like electron donating and electron withdrawing groups in quinoline and benzene were diversely responsible for a broad range of antimicrobial, antituberculosis, antimalarial and antioxidant activities. In the benzene ring electron donating group like $-\text{CH}_3$ and $-\text{OCH}_3$ exhibited decrease in the antibacterial activities, whereas the similar group showed increased activity in case of antimalarial, antituberculosis as well as in antioxidant activities. Also compounds having electron withdrawing group in benzene ring showed excellent antibacterial activity against *S. Pneumoniae*. On the other side, quinoline ring containing $-\text{CH}_3$ showed very good antimalarial, antituberculosis activity as well as showed best antibacterial activity against all 3 gram positive bacteria. In 1,3,4-oxadiazole nucleus, the ring containing nitrogen at 3rd or 4th position showed excellent antimalarial, antitubercular, antifungal, antioxidant as well as antibacterial activities.

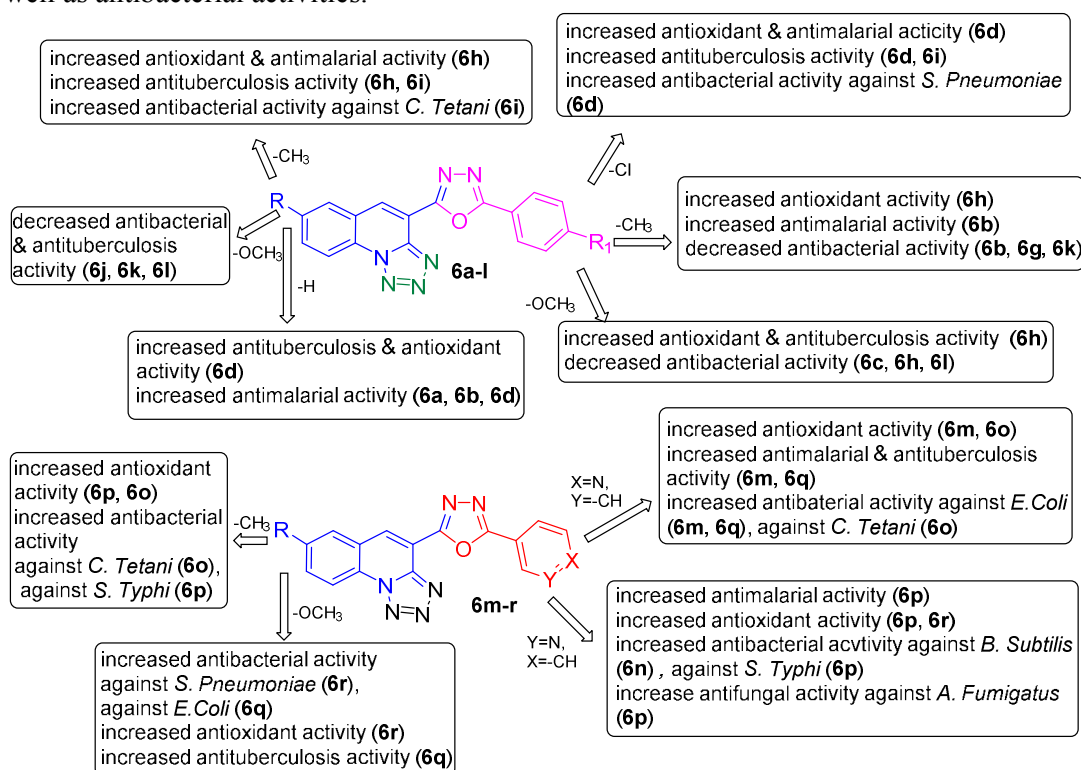


Fig.1. Structure-activity relationship for antimicrobial, antituberculosis, antimalarial and antioxidant activity of the synthesized compound 6a-r.

3. Conclusion

This study presents the synthesis of 18 new quinoline – oxadiazole hybrids which showed some brilliant biological results. This work helps to corroborate the choice of the quinoline based 1,3,4-oxadiazole scaffold as a useful model for designing new antimicrobial, antitubercular, antimalarial and antioxidant agents. This synthetic approach allows the addition of potent bioactive nuclei in a single scaffold in an easy way. Two leading candidates (6h and 6q) displayed modest antituberculosis activity. Some of the compounds showed brilliant antimalarial activity against *P. falciparum* strains as compared to quinine.

Compound **6q** has emerged as the most promising antimicrobial member of the series, showing better antitubercular, antimalarial as well antioxidant activities. The results indicated that oxadiazoles scaffold which is clubbed with the pyridine ring showed the highest antibacterial, antituberculosis and antimalarial activities.

4. Experimental section

4.1. Chemistry

All the chemicals were purchased from Sigma Aldrich, Spectrochem and Merck – India. Commercial grade solvents were used and were distilled before use. Melting points were determined using a melting point apparatus μ ThermoCal₁₀ (Analab Scientific Pvt. Ltd, India) and are uncorrected. The completion of the reactions was checked by thin-layer chromatography (TLC) on aluminium plates coated with silica gel 60 F₂₅₄, 0.25 mm thickness (Merck) and detection of the components was made by exposure to iodine vapours or UV light. The IR spectra (in KBr pellets) were recorded on a Perkin-Elmer Spectrum GX FT-IR 157 spectrophotometer (Perkin-Elmer, USA) and only the characteristic peaks are reported in cm^{-1} . ¹H-NMR spectra were recorded in DMSO-*d*₆ on a Bruker Avance 400F (MHz) spectrometer (Bruker Scientific Corporation Ltd., Switzerland) using the residual solvent signal as an internal standard at 400 MHz. Chemical shifts (δ) are given in ppm and coupling constants (*J*) are in Hz. Mass spectra were recorded on a Shimadzu LCMS 2010 spectrometer (Shimadzu, Tokyo, Japan) at Sardar Patel University (PURSE programme of DST), Vallabh Vidyanagar. The elemental analysis was carried out by using a Perkin-Elmer 2400 series-II elemental analyzer (Perkin-Elmer, USA) and all compounds were found to be within $\pm 0.4\%$ of the theoretical compositions.

4.2. General procedure for the synthesis of 7-substituted tetrazolo[1,5-*a*]quinoline-4-carbaldehyde (**3a-c**).

2-chloroquinoline-3-carbaldehyde **1** (1 mmol) and sodium azide **2** (2 mmol) in the presence of catalytic amount of acetic acid and ethanol (10 ml) were charged in a 100 mL round bottom flask equipped with a mechanical stirrer and a condenser. The reaction mixture was refluxed at 78 °C for 2-3 hr. The progress of the reaction was monitored by TLC. After the completion of reaction as confirmed by TLC, the separated crude product was filtered, washed thoroughly with ethanol, dried and recrystallized from chloroform to obtain a yellowish-white solid (**3a-c**).

4.3. General procedure for the synthesis of (*E*)-4-substituted-N'-((7-substituted tetrazolo [1,5-*a*]quinolin-4-yl)methylene)benzohydrazide (**5a-l**).

A mixture of 7-substituted tetrazolo[1,5-*a*]quinoline-4-carbaldehyde (**3a-c**) (10 mmol), 4-substituted benzohydrazide **4a-e** (10 mmol) and catalytic amount of glacial acetic acid in methanol (40 mL) was refluxed for 1 h. After the completion of reaction, the reaction mixture was stirred magnetically for further 10-15 min. After cooling the separated solid mass was collected by filtration, washed well with ethanol (10 mL) dried, and crystallized from hot ethanol (10 mL) to affording compounds (**5a-l**).

4.4. General procedure for the synthesis of (*E*)-N'-((7-substituted tetrazolo [1,5-*a*]quinolin-4-yl)methylene)nicotinohydrazide/isonicotinohydrazide (**5m-r**).

A mixture of 7-substituted tetrazolo[1,5-*a*]quinoline-4-carbaldehyde (**3a-c**) (10 mmol), nicotinohydrazide (**4a'**) and/or isonicotinohydrazide (**4b'**) (10 mmol) and catalytic amount of glacial acetic acid in methanol (40 mL) was refluxed for 1 h. After the completion of

reaction, the reaction mixture was stirred magnetically for further 10-15 min. After cooling the separated solid mass was collected by filtration, washed well with ethanol (10 mL) dried, and crystallized from hot ethanol (10 mL) to affording compounds (**5m-r**).

4.5. General procedure for the synthesis of 2-(7-substituted tetrazolo [1,5-a]quinolin-4-yl)-5-(p-substituted)-1,3,4-oxadiazole (**6a-l**).

A mixture of compound **5a-l** (10 mmol) was dissolved in DCM (20 ml) and continuously stirred. To this solution, $\text{PhI}(\text{OAc})_2$ (10 mmol) was added and the mixture was stirred for 15-20 min at room temperature. After the completion of the reaction as monitored by TLC (ethyl acetate: hexane: 3:7), the solvent was evaporated and the residue was washed with diethyl ether, filtered (5 mL), dried and then crystallized from acetone to affording target compounds (**6a-l**).

4.5.1. 2-phenyl-5-(tetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (**6a**)

White solid, Yield 93 %; m.p. 213-215 °C; FTIR (KBr, ν_{max} , cm^{-1}): 1623 and 1592 (C=N and C=C); 3052 (Ar, -CH str.); ^1H NMR (400 MHz, $\text{DMSO-}d_6$): δ 7.57 -8.75 (m, 9H, Ar-H), 9.30 (s, 1H, Ar-H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ : 116.7, 126.9, 127.5, 128.4, 128.8, 128.9, 129.3, 130.1, 130.7, 132.9, 135.2, 146.0, 149.2, 164.1; ESI-MS (m/z): 314.31 (M+); Anal. Calcd (%) for $\text{C}_{17}\text{H}_{10}\text{N}_6\text{O}$: C, 64.96; H, 3.21; N, 26.74; Found: C, 64.93; H, 3.25; N, 26.77.

4.5.2. 2-(tetrazolo[1,5-a]quinolin-4-yl)-5-(p-tolyl)-1,3,4-oxadiazole (**6b**)

Yellowish solid, Yield 73 %; m.p. 207-209 °C; FTIR (KBr, ν_{max} , cm^{-1}): 1622 and 1592 (C=N and C=C); 3053 (Ar, -CH str.); ^1H NMR (400 MHz, $\text{DMSO-}d_6$): δ 2.46 (s, 3H, $-\text{CH}_3$ of benzene ring), 7.51 -8.74 (m, 8H, Ar-H), 9.27 (s, 1H, Ar-H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ : 21.2, 116.7, 126.4, 127.1, 127.7, 128.6, 128.8, 130.3, 131.9, 136.0, 142.5, 146.4, 149.8, 165.6, 165.7; ESI-MS (m/z): 328.34 (M+); Anal. Calcd (%) for $\text{C}_{18}\text{H}_{12}\text{N}_6\text{O}$: C, 65.85; H, 3.68; N, 25.60; Found: C, 65.82; H, 3.66; N, 25.64.

4.5.3. 2-(4-methoxyphenyl)-5-(tetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (**6c**)

White solid, Yield 81 %; m.p. 198-200 °C; FTIR (KBr, ν_{max} , cm^{-1}): 1625 and 1598 (C=N and C=C); 3057 (Ar, -CH str.); ^1H NMR (400 MHz, $\text{DMSO-}d_6$): δ 3.91 (s, 3H, $-\text{OCH}_3$ of benzene ring), 7.57 -8.74 (m, 8H, Ar-H), 9.26 (s, 1H, Ar-H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ : 54.2, 114.3, 115.5, 116.0, 126.8, 127.5, 128.7, 129.0, 129.8, 130.8, 135.6, 146.0, 149.8, 160.3, 165.2; ESI-MS (m/z): 344.33 (M+); Anal. Calcd (%) for $\text{C}_{18}\text{H}_{12}\text{N}_6\text{O}_2$: C, 62.79; H, 3.51; N, 24.41; Found: C, 62.77; H, 3.49; N, 24.43.

4.5.4. 2-(4-chlorophenyl)-5-(tetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (**6d**)

Yellowish white solid, Yield 80 %; m.p. 196-198 °C; FTIR (KBr, ν_{max} , cm^{-1}): 1638 and 1593 (C=N and C=C); 3055 (Ar, -CH str.); ^1H NMR (400 MHz, $\text{DMSO-}d_6$): δ 7.53 -8.75 (m, 8H, Ar-H), 9.21 (s, 1H, Ar-H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ : 116.7, 124.6, 127.3, 128.5, 128.8, 128.9, 129.2, 130.1, 130.7, 134.3, 135.3, 145.8, 149.7, 164.9; ESI-MS (m/z): 348.75 (M+), 350.81 (M+2); Anal. Calcd (%) for $\text{C}_{17}\text{H}_9\text{ClN}_6\text{O}$: C, 58.55; H, 2.60; N, 24.10; Found: C, 58.53; H, 2.61; N, 24.14.

4.5.5. 2-(4-bromophenyl)-5-(tetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (**6e**)

Yellow solid, Yield 86 %; m.p. 191-193 °C; FTIR (KBr, ν_{max} , cm^{-1}): 1631 and 1598 (C=N and C=C); 3067 (Ar, -CH str.); ^1H NMR (400 MHz, $\text{DMSO-}d_6$): δ 7.52 -9.18 (m, 9H, Ar-H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ : 116.7, 123.3, 125.7, 126.9, 128.1, 128.5, 129.2, 129.8,

130.7, 132.7, 135.6, 145.4, 149.7, 165.9; ESI-MS (m/z): 393.20 (M⁺), 395.13 (M+2); Anal. Calcd (%) for C₁₇H₉BrN₆O: C, 51.93; H, 2.31; N, 21.37; Found: C, 51.91; H, 2.29; N, 21.33.

4.5.6. 2-(7-methyltetrazolo[1,5-a]quinolin-4-yl)-5-phenyl-1,3,4-oxadiazole (6f)

White solid, Yield 83 %; m.p. 238-240 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1624 and 1590 (C=N and C=C); 3053(Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.33 (s, 3H, -CH₃ of quinoline ring), 7.55 -8.75 (m, 8H, Ar-H), 9.31 (s, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.5, 126.9, 127.1, 128.4, 128.6, 129.5, 131.6, 130.9, 133.6, 134.7, 136.3, 141.8, 149.7, 164.8; ESI-MS (m/z): 328.34 (M⁺); Anal. Calcd (%) for C₁₈H₁₂N₆O: C, 65.85; H, 3.68; N, 25.60; Found: C, 65.83; H, 3.70; N, 25.64.

4.5.7. 2-(7-methyltetrazolo[1,5-a]quinolin-4-yl)-5-(p-tolyl)-1,3,4-oxadiazole (6g)

White solid, Yield 79 %; m.p. 197-199 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1627 and 1592 (C=N and C=C); 3056(Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.33 (s, 3H, -CH₃ of quinoline ring), 2.67 (s, 3H, -CH₃ of benzene ring), 7.54 -8.64 (m, 7H, Ar-H), 9.19 (s, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.4, 21.9, 126.4, 127.5, 127.9, 128.6, 128.9, 130.2, 131.3, 131.9, 134.6, 136.8, 142.8, 144.1, 149.3, 164.9; ESI-MS (m/z): 342.36 (M⁺); Anal. Calcd (%) for C₁₉H₁₄N₆O: C, 66.66; H, 4.12; N, 24.55; Found: C, 66.63; H, 4.14; N, 24.53.

4.5.8. 2-(4-methoxyphenyl)-5-(7-methyltetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (6h)

Yellowish white, Yield 74 %; m.p. 201-203 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1622 and 1594 (C=N and C=C); 3060(Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.33 (s, 3H, -CH₃ of quinoline ring), 3.92 (s, 3H, -OCH₃ of benzene ring), 7.53 -9.22 (m, 8H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.8, 55.9, 114.3, 115.8, 126.7, 128.5, 128.9, 129.4, 130.8, 131.8, 134.6, 136.8, 144.0, 148.6, 160.3, 165.5; ESI-MS (m/z): 358.36 (M⁺); Anal. Calcd (%) for C₁₉H₁₄N₆O₂: C, 63.68; H, 3.94; N, 23.45; Found: C, 63.66; H, 3.96; N, 23.48.

4.5.9. 2-(4-chlorophenyl)-5-(7-methyltetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (6i)

White solid, Yield 89 %; m.p. 233-235 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1628 and 1593 (C=N and C=C); 3053(Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.33 (s, 3H, -CH₃ of quinoline ring), 7.55 -9.23 (m, 8H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.7, 124.3, 126.9, 128.4, 128.7, 128.9, 129.8, 130.5, 131.7, 134.6, 135.1, 136.7, 143.8, 148.9, 165.7; ESI-MS (m/z): 362.78 (M⁺), 364.65 (M+2); Anal. Calcd (%) for C₁₈H₁₁ClN₆O: C, 59.59; H, 3.06; N, 23.17; Found: C, 59.60; H, 3.09; N, 23.16.

4.5.10. 2-(7-methoxytetrazolo[1,5-a]quinolin-4-yl)-5-phenyl-1,3,4-oxadiazole (6j)

White solid, Yield 76 %; m.p. 220-222 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1625 and 1591 (C=N and C=C); 3062 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 3.90 (s, 3H, -OCH₃ of quinoline ring), 7.56 -8.73 (m, 8H, Ar-H), 9.27 (s, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 55.4, 103.9, 120.0, 127.5, 127.8, 129.2, 129.5, 130.9, 131.2, 133.6, 133.9, 141.7, 146.8, 157.1, 165.2; ESI-MS (m/z): 344.33 (M⁺); Anal. Calcd (%) for C₁₈H₁₂N₆O₂: C, 62.79; H, 3.51; N, 24.41; Found: C, 62.77; H, 3.49; N, 24.43.

4.5.11. 2-(7-methoxytetrazolo[1,5-a]quinolin-4-yl)-5-(p-tolyl)-1,3,4-oxadiazole (6k)

Yellowish white, Yield 84 %; m.p. 223-225 °C ; FTIR (KBr, ν_{\max} , cm⁻¹): 1623 and 1591 (C=N and C=C); 3067 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.46 (s, 3H, -CH₃ of benzene ring), 3.99 (s, 3H, -OCH₃ of quinoline ring), 7.52 -8.66 (m, 8H, Ar-H), 9.19 (s, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.8, 55.7, 105.9, 122.8, 126.7, 127.6, 129.8,

130.8, 131.2, 131.9, 134.6, 141.5, 142.8, 147.2, 157.8, 164.9; ESI-MS (m/z): 358.36 (M⁺); Anal. Calcd (%) for C₁₉H₁₄N₆O₂: C, 63.68; H, 3.94; N, 23.45; Found: C, 63.71; H, 3.92; N, 23.46.

4.5.12. 2-(4-methoxyphenyl)-5-(7-methoxytetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (6l)

White solid, Yield 80 %; m.p. 208-210 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1625 and 1593 (C=N and C=C); 3052 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 3.89 (s, 3H, -OCH₃ of benzene ring), 3.98 (s, 3H, -OCH₃ of quinoline ring), 7.54 -9.23 (m, 8H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 55.6, 55.9, 105.8, 114.8, 115.3, 122.7, 129.1, 129.8, 130.7, 131.5, 134.6, 141.8, 147.3, 157.4, 160.7, 165.2; ESI-MS (m/z): 374.36 (M⁺); Anal. Calcd (%) for C₁₉H₁₄N₆O₃: C, 60.96; H, 3.77; N, 22.45; Found: C, 60.99; H, 3.78; N, 22.43.

4.6. General procedure for the synthesis of 2-(7-substitutedtetrazolo [1,5-a]quinolin-4-yl)-5-(pyridin-3/4-yl)-1,3,4-oxadiazole (6m-r).

A mixture of compound **5m-r** (10 mmol) was dissolved in DCM (20 ml) and continuously stirred. To this solution, PhI(OAc)₂ (10 mmol) was added and the mixture was stirred for 15-20 min at room temperature. After the completion of the reaction as monitored by TLC (ethyl acetate: hexane: 5:5), the solvent was evaporated and the residue was washed with diethyl ether, filtered (5 mL), dried and then crystallized from acetone to affording target compounds (**6m-r**).

4.6.1. 2-(pyridin-4-yl)-5-(tetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (6m)

Light yellow solid, Yield 84 %; m.p. 201-203 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1622 and 1590 (C=N and C=C); 3052 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.84 -8.83 (m, 9H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 116.7, 122.1, 124.3, 128.9, 130.6, 131.0, 132.6, 144.8, 147.7, 150.2, 164.1, 165.5; ESI-MS (m/z): 315.30 (M⁺); Anal. Calcd (%) for C₁₆H₉N₇O: C, 60.95; H, 2.88; N, 31.10; Found: C, 60.92; H, 2.87; N, 31.13.

4.6.2. 2-(pyridin-3-yl)-5-(tetrazolo[1,5-a]quinolin-4-yl)-1,3,4-oxadiazole (6n)

White solid, Yield 77 %; m.p. 215-217 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1634 and 1589 (C=N and C=C); 3051 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.85 -8.89 (m, 9H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 116.7, 122.1, 124.3, 128.9, 130.6, 131.0, 132.6, 144.8, 147.7, 150.2, 164.1, 165.5; ESI-MS (m/z): 315.30 (M⁺); Anal. Calcd (%) for C₁₆H₉N₇O: C, 60.95; H, 2.88; N, 31.10; Found: C, 60.93; H, 2.90; N, 31.14.

4.6.3. 2-(7-methyltetrazolo[1,5-a]quinolin-4-yl)-5-(pyridin-4-yl)-1,3,4-oxadiazole (6o)

Yellowish white, Yield 86 %; m.p. 200-202 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1632 and 1590 (C=N and C=C); 3053 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.55 (s, 3H, -CH₃ of quinoline ring), 7.77 -8.96 (m, 8H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.5, 121.1, 126.9, 127.9, 128.4, 130.7, 131.2, 134.4, 136.9, 143.2, 143.7, 148.6, 149.9, 164.8; ESI-MS (m/z): 329.32 (M⁺); Anal. Calcd (%) for C₁₇H₁₁N₇O: C, 62.00; H, 3.37; N, 29.77; Found: C, 62.04; H, 3.41; N, 29.75.

4.6.4. 2-(7-methyltetrazolo[1,5-a]quinolin-4-yl)-5-(pyridin-3-yl)-1,3,4-oxadiazole (6p)

White solid, Yield 81 %; m.p. 216-218 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1628 and 1592 (C=N and C=C); 3055 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 2.55 (s, 3H, -CH₃ of quinoline ring), 7.75 -8.94 (m, 8H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 21.8, 124.0, 124.7, 126.8, 128.4, 128.9, 130.8, 131.6, 134.0, 134.8, 136.7, 144.1, 147.3, 148.8, 152.5, 165.6;

ESI-MS (m/z): 329.32 (M⁺); Anal. Calcd (%) for C₁₇H₁₁N₇O: C, 62.00; H, 3.37; N, 29.77; Found: C, 62.03; H, 3.39; N, 29.79.

4.6.5. 2-(7-methoxytetrazolo[1,5-a]quinolin-4-yl)-5-(pyridin-4-yl)-1,3,4-oxadiazole (6g)

Yellow solid, Yield 85 %; m.p. 197-199 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1622 and 1592(C=N and C=C); 3056 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 3.90 (s, 3H, -OCH₃ of quinoline ring), 7.45 -8.48 (m, 7H, Ar-H), 8.76 (s, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ : 55.7, 105.5, 121.8, 122.9, 130.1, 131.8, 134.6, 141.5, 143.8, 147.7, 150.0, 157.8, 164.9; ESI-MS (m/z): 345.32 (M⁺); Anal. Calcd (%) for C₁₇H₁₁N₇O₂: : C, 59.13; H, 3.21; N, 28.39; Found: C, 59.11 H, 3.19; N, 28.42.

4.6.6. 2-(7-methoxytetrazolo[1,5-a]quinolin-4-yl)-5-(pyridin-3-yl)-1,3,4-oxadiazole (6r)

White solid, Yield 83 %; m.p. 220-222 °C; FTIR (KBr, ν_{\max} , cm⁻¹): 1620 and 1592(C=N and C=C); 3053 (Ar, -CH str.); ¹H NMR (400 MHz, DMSO-*d*₆): δ 3.91 (s, 3H, -OCH₃ of quinoline ring), 7.48 -8.47 (m, 7H, Ar-H), 8.80 (s, 1H, Ar-H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ : 55.8, 105.9, 122.7, 124.0, 124.8, 129.7, 130.6, 131.3, 134.3, 134.8, 141.9, 147.7, 148.0, 152.8, 157.4, 164.8; ESI-MS (m/z): 345.32 (M⁺); Anal. Calcd (%) for C₁₇H₁₁N₇O₂: C, 59.13; H, 3.21; N, 28.39; Found: C, 59.15; H, 3.23; N, 28.37.

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References

- [I] A. Matin, N. Gavande, M.S. Kim, N.X. Yang, N.K. Salam, J.R. Hanrahan, R.H. Roubin, D.E. Hibbs, 7-Hydroxy-benzopyran-4-one derivatives: a novel pharmacophore of peroxisome proliferator-activated receptor α and γ (PPAR α and γ) dual agonists, Journal of medicinal chemistry, 52 (2009) 6835-6850.
- [II] D.H. Boschelli, D.T. Connor, D.A. Bornemeier, R.D. Dyer, J.A. Kennedy, P.J. Kuipers, G.C. Okonkwo, D.J. Schrier, C.D. Wright, 1, 3, 4-Oxadiazole, 1, 3, 4-thiadiazole, and 1, 2, 4-triazole analogs of the fenamates: in vitro inhibition of cyclooxygenase and 5-lipoxygenase activities, Journal of medicinal chemistry, 36 (1993) 1802-1810.
- [III] C.B. Chapleo, P.L. Myers, A.C. Smith, M.R. Stillings, I.F. Tulloch, D.S. Walter, Substituted 1, 3, 4-thiadiazoles with anticonvulsant activity. 4. Amidines, Journal of medicinal chemistry, 31 (1988) 7-11.
- [IV] A.A. El-Emam, O.A. Al-Deeb, M. Al-Omar, J. Lehmann, Synthesis, antimicrobial, and anti-HIV-1 activity of certain 5-(1-adamantyl)-2-substituted thio-1, 3, 4-oxadiazoles and 5-(1-adamantyl)-3-substituted aminomethyl-1, 3, 4-oxadiazoline-2-thiones, Bioorganic & medicinal chemistry, 12 (2004) 5107-5113.
- [V] Y. Hu, C.-Y. Li, X.-M. Wang, Y.-H. Yang, H.-L. Zhu, 1, 3, 4-Thiadiazole: synthesis, reactions, and applications in medicinal, agricultural, and materials chemistry, Chemical reviews, 114 (2014) 5572-5610.

- [VI] S. Ningaiah, U.K. Bhadraiah, S.D. Doddaramappa, S. Keshavamurthy, C. Javarasetty, Novel pyrazole integrated 1, 3, 4-oxadiazoles: Synthesis, characterization and antimicrobial evaluation, *Bioorganic & medicinal chemistry letters*, 24 (2014) 245-248.
- [VII] M.-Y. Yang, W. Zhao, Z.-H. Sun, C.-X. Tan, J.-Q. Weng, X.-H. Liu, Synthesis and biological activity of acylthiourea derivatives contain 1, 2, 3-thiadiazole and 1, 3, 4-thiadiazole, *Letters in Drug Design & Discovery*, 12 (2015) 314-318.
- [VIII] W.H. Organization, Global tuberculosis report 2015, World Health Organization, 2015.
- [IX] M. Arentz, T.R. Hawn, Tuberculosis infection: insight from immunogenomics, *Drug Discovery Today: Disease Mechanisms*, 4 (2008) 231-236.
- [X] Global Tuberculosis Controlesurveillance, Planning, Financing. WHO Report (2006).http://www.who.int/tb/publications/global_report/2006/pdf/full-report_correctedversion.pdf.
- [XI] M.S. Yar, A.A. Siddiqui, M. Ashraf Ali, Synthesis and Anti Tuberculostatic Activity of Novel 1, 3, 4-Oxadiazole Derivatives, *Journal of the Chinese Chemical Society*, 54 (2007) 5-8.
- [XII] A. Nikalje, P. Mudassar, Multidrug-resistant Mycobacterium tuberculosis: A brief review, *Asian J. Biol. Sci*, 4 (2011).
- [XIII] W.H. Organization, World malaria report 2014. 2014, Avarilable: http://www.who.int/malaria/publications/world_malaria_report_2014/en/. Accessed, 24 (2015).
- [XIV] A.M. Dondorp, F. Nosten, P. Yi, D. Das, A.P. Phyo, J. Tarning, K.M. Lwin, F. Ariey, W. Hanpithakpong, S.J. Lee, Artemisinin resistance in Plasmodium falciparum malaria, *New England Journal of Medicine*, 361 (2009) 455-467.
- [XV] L. Ursos, P.D. Roepe, Chloroquine resistance in the malarial parasite, Plasmodium falciparum, *Medicinal research reviews*, 22 (2002) 465-491.
- [XVI] G.D. Henry, De novo synthesis of substituted pyridines, *Tetrahedron*, 60 (2004) 6043-6061.
- [XVII] P. M Njogu, K. Chibale, Recent developments in rationally designed multitarget antiprotozoan agents, *Current medicinal chemistry*, 20 (2013) 1715-1742.
- [XVIII] T. Shigeyama, K. Katakawa, N. Kogure, M. Kitajima, H. Takayama, Asymmetric total syntheses of two phlegmarine-type alkaloids, lycoposerramines-V and-W, newly isolated from *Lycopodium serratum*, *Organic letters*, 9 (2007) 4069-4072.
- [XIX] A. Shirai, O. Miyata, N. Tohnai, M. Miyata, D.J. Procter, D. Sucunza, T. Naito, Total Synthesis of (-)-Martinellie Acid via Radical Addition– Cyclization– Elimination Reaction, *The Journal of organic chemistry*, 73 (2008) 4464-4475.
- [XX] A. Andreani, M. Granaiola, A. Leoni, A. Locatelli, R. Morigi, M. Rambaldi, Synthesis and antitubercular activity of imidazo [2, 1-b] thiazoles, *European journal of medicinal chemistry*, 36 (2001) 743-746.
- [XXI] N.N. Farshori, M.R. Bandy, A. Ahmad, A.U. Khan, A. Rauf, Synthesis, characterization, and in vitro antimicrobial activities of 5-alkenyl/hydroxyalkenyl-2-phenylamine-1, 3, 4-oxadiazoles and thiadiazoles, *Bioorganic & medicinal chemistry letters*, 20 (2010) 1933-1938.
- [XXII] P. Li, L. Shi, X. Yang, L. Yang, X.-W. Chen, F. Wu, Q.-C. Shi, W.-M. Xu, M. He, D.-Y. Hu, Design, synthesis, and antibacterial activity against rice bacterial leaf blight and leaf streak of 2, 5-substituted-1, 3, 4-oxadiazole/thiadiazole sulfone derivative, *Bioorganic & medicinal chemistry letters*, 24 (2014) 1677-1680.

- [XXIII] N.B. Patel, A.C. Purohit, D.P. Rajani, R. Moo-Puc, G. Rivera, New 2-benzylsulfanyl-nicotinic acid based 1, 3, 4-oxadiazoles: Their synthesis and biological evaluation, *European journal of medicinal chemistry*, 62 (2013) 677-687.
- [XXIV] R.A. Rane, P. Bangalore, S.D. Borhade, P.K. Khandare, Synthesis and evaluation of novel 4-nitropyrrole-based 1, 3, 4-oxadiazole derivatives as antimicrobial and anti-tubercular agents, *European journal of medicinal chemistry*, 70 (2013) 49-58.
- [XXV] S. Bondock, S. Adel, H.A. Etman, F.A. Badria, Synthesis and antitumor evaluation of some new 1, 3, 4-oxadiazole-based heterocycles, *European journal of medicinal chemistry*, 48 (2012) 192-199.
- [XXVI] A.T. Taher, H.H. Georgey, H.I. El-Subbagh, Novel 1, 3, 4-heterodiazole analogues: Synthesis and in-vitro antitumor activity, *European journal of medicinal chemistry*, 47 (2012) 445-451.
- [XXVII] J. Bergman, A. Brynolf, Synthesis of Chrysogine, a Metabolite of *Penicillium chrysogenum* and some related 2-substituted 4-(3H)-Quinazolinones, *Tetrahedron*, 46 (1990) 1295-1310.
- [XXVIII] O. Prakash, M. Kumar, R. Kumar, C. Sharma, K. Aneja, Hypervalent iodine (III) mediated synthesis of novel unsymmetrical 2, 5-disubstituted 1, 3, 4-oxadiazoles as antibacterial and antifungal agents, *European journal of medicinal chemistry*, 45 (2010) 4252-4257.
- [XXIX] S. Bansal, M. Bala, S.K. Suthar, S. Choudhary, S. Bhattacharya, V. Bhardwaj, S. Singla, A. Joseph, Design and synthesis of novel 2-phenyl-5-(1, 3-diphenyl-1H-pyrazol-4-yl)-1, 3, 4-oxadiazoles as selective COX-2 inhibitors with potent anti-inflammatory activity, *European journal of medicinal chemistry*, 80 (2014) 167-174.
- [XXX] R.D. Dillard, D.E. Pavey, D.N. Benslay, Synthesis and antiinflammatory activity of some 2, 2-dimethyl-1, 2-dihydroquinolines, *Journal of medicinal chemistry*, 16 (1973) 251-253.
- [XXXI] C. Fattorusso, G. Campiani, G. Kukreja, M. Persico, S. Butini, M.P. Romano, M. Altarelli, S. Ros, M. Brindisi, L. Savini, Design, synthesis, and structure-activity relationship studies of 4-quinolinyl- and 9-acrydinyldiazones as potent antimalarial agents, *Journal of medicinal chemistry*, 51 (2008) 1333-1343.
- [XXXII] S. Manohar, M. Tripathi, D. S Rawat, 4-Aminoquinoline Based Molecular Hybrids as Antimalarials: An Overview, *Current topics in medicinal chemistry*, 14 (2014) 1706-1733.
- [XXXIII] F.W. Muregi, A. Ishih, Next-generation antimalarial drugs: hybrid molecules as a new strategy in drug design, *Drug development research*, 71 (2010) 20-32.
- [XXXIV] S. Vandekerckhove, M. D'hooghe, Quinoline-based antimalarial hybrid compounds, *Bioorganic & medicinal chemistry*, 23 (2015) 5098-5119.
- [XXXV] M. Ogata, H. ATOBE, H. KUSHIDA, K. YAMAMOTO, In vitro sensitivity of mycoplasmas isolated from various animals and sewage to antibiotics and nitrofurans, *The Journal of antibiotics*, 24 (1971) 443-451.
- [XXXVI] R. Laufer, O.G. Paz, A. Di Marco, F. Bonelli, E. Monteagudo, V. Summa, M. Rowley, Quantitative prediction of human clearance guiding the development of raltegravir (MK-0518, Isentress) and related HIV integrase inhibitors, *Drug Metabolism and Disposition*, 37 (2009) 873-883.
- [XXXVII] O. Meth-Cohn, B. Narine, A versatile new synthesis of quinolines, thienopyridines and related fused pyridines, *Tetrahedron Letters*, 19 (1978) 2045-2048.
- [XXXVIII] P. Wayne, National committee for clinical laboratory standards, Performance standards for antimicrobial disc susceptibility testing, 12 (2002).

- [XXXIX] H.G. Kathrotiya, M.P. Patel, Synthesis and identification of β -aryloxyquinoline based diversely fluorine substituted N-aryl quinolone derivatives as a new class of antimicrobial, antituberculosis and antioxidant agents, *European journal of medicinal chemistry*, 63 (2013) 675-684.
- [XL] H.H. Jardosh, M.P. Patel, Design and synthesis of biquinolone–isoniazid hybrids as a new class of antitubercular and antimicrobial agents, *European journal of medicinal chemistry*, 65 (2013) 348-359.
- [XLI] A. Rattan, *Antimicrobials in laboratory medicine*, Churchill BI, Livingstone, New Delhi, 85 (2000).
- [XLII] I.F. Benzie, J. Strain, The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay, *Analytical biochemistry*, 239 (1996) 70-76.

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